









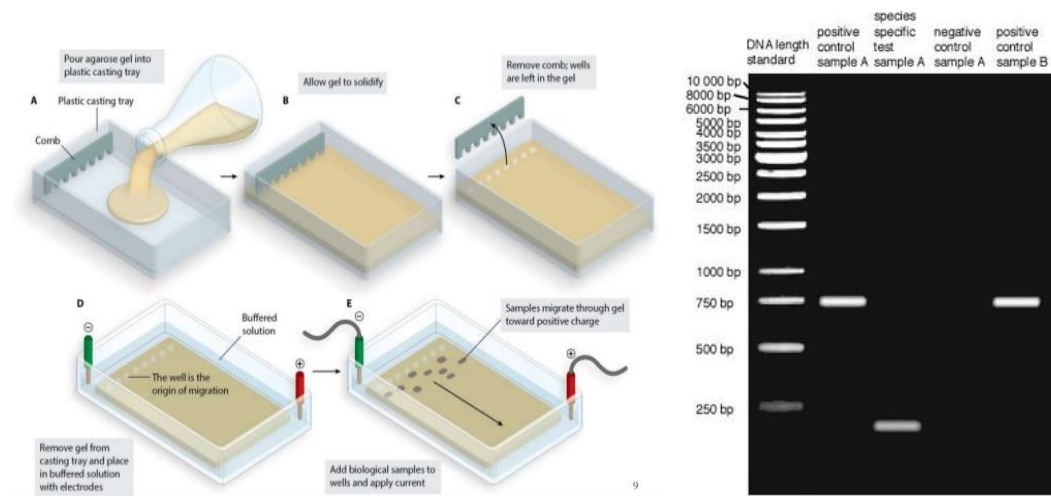


## **Agarose Gel Electrophoresis**

When significant quantity of DNA was produced during the PCR reaction, they can be analyzed through gene electrophoresis. This technique uses a carbohydrate  is derived from seaweeds. Agarose is mixed with  Tris-EDTA-acetate (TAE) or Tris-EDTA-borate (TBE)  til it melts and mixes with the buffer. The resulting  es is then casted on the agarose gel electrophoresis  d is allowed to polymerize. Polymerization solution  el . The agarose gel contains wells where samples  s portion of the gel where the wells are located  egative electrode of the electrophoresis apparatus  ode on the other side is positively charged. The  ed into the same buffer that was used for its preparation. DNA samples are placed into the wells. The net charge of the DNA is negative due to its phosphate backbone, such that when the current is allowed to pass through the agarose gel, the negatively-charged DNA molecules will be attracted to move towards the positively-charged electrode. Larger DNA fragments will migrate through the gel slowly, while shorter and lighter DNA amplicons will move faster across the gel. In order to know how many bases are contained by the amplicon, molecular weight markers are used in every electrophoresis experiment. The migration of the DNA sample within the gel is monitored through a gel leading

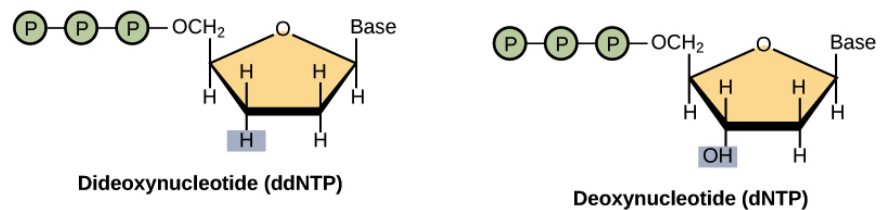
dye that is mixed with the DNA. The reactions are colorless solutions. After separation according to their molecular sizes, the agarose gel is visualized using gel documentation equipment. In most cases, staining agents are added in the agarose gel. The DNA fragments themselves into the double helix DNA and exhibit fluorescence when struck by UV radiation. The more intense the fluorescence, the higher the concentration of DNA has been generated by the PCR reaction. Furthermore, agarose gel electrophoresis can also be used to purify DNA fragments before they are analyzed through DNA sequencing.



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<https://image.slidesharecdn.com/474f3676-3154-4b76-b3d0-6a082debe892-160718131054/95/gel-electrophoresis-9-638.jpg?cb=1468847472>

## 9.6 Sanger-Coulson Method for DNA Sequencing

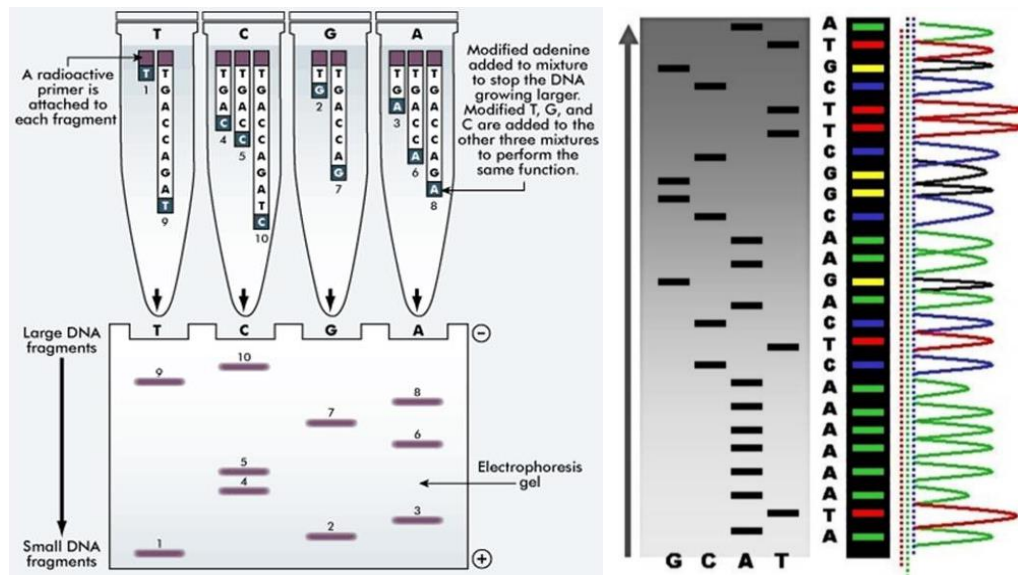
After gel electrophoresis, the nucleotide sequence of the amplicons are determined. One of the methods in determination of a DNA sequence is the Sanger method. The amplicon is aliquoted into four separate tubes, each amplified through PCR using the same primers and the same amplicon. Each reaction tube contains a specific dideoxynucleotide or ddNTPs (i.e., ddATP, ddGTP, ddCTP and ddTTP) as shown in Figure X, the ddNTP lacks oxygen atom at the 3' carbon of the sugar. This prevents attachment of the next nucleotide and is not incorporated in the growing DNA sequence by the DNA polymerase.



Source: <https://ka-perseus->

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As a result, replication stops on every position where the ddNTP are incorporated, leading to formation of DNA amplicons with varied molecular weights in four separate tubes. The amplicons are run in an agarose or a capillary gel for separation. Results are read from the bottom part of the gel where the DNA fragments are shortest. Nowadays, results of DNA sequencing, called chromatogram are read using computer programs,



Source:[http://mol-biol4masters.masters.grkraj.org/html/DNA\\_C\\_Value\\_Paradox2-Hybridization\\_Kinetics.htm](http://mol-biol4masters.masters.grkraj.org/html/DNA_C_Value_Paradox2-Hybridization_Kinetics.htm)

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especially for very long DNA sequences. Through these basic concepts of gene manipulation, genetic engineers are capable of producing organisms with various types as well as with other applications that address the agricultural, medical, educational and industrial development of pest and drought resistant and high yielding insects for pest control, glowing bacteria that can be used in absence of microscopes, larger-than-usual fruits to address global hunger, medicines specifically designed for various diseases, and many other products and commodities are generated through alteration of gene structure and function of organisms.